

TRANSLATION NO. 99

DATE: Sight 1968

DDC AVAILABILITY NOTICE

This document has been approved for public release and sale; its distribution is unlimited.

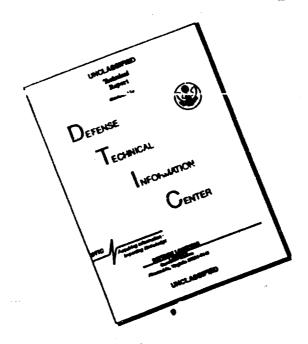
N a

DEPARTMENT OF THE ARMY Fort Detrick Frederick, Maryland

Record and by the CLEARINGHOUSE for Federal Scientific & Technical Information Springfield Va. 22151

1

ISCLAIMER NOTICE



THIS DOCUMENT IS BEST QUALITY AVAILABLE. THE COPY FURNISHED TO DTIC CONTAINED A SIGNIFICANT NUMBER OF PAGES WHICH DO NOT REPRODUCE LEGIBLY.

Trans. Acad. Sci., USSE., 1955, v. 105(2) Hov; 315-318.

On the chemical composition of the antigonic substances of the agent of tuleremia.

In this report the virmlent strain, No. 21, of the Easterium tulerence, representing the S-form of microbe, was utilized. The cultures were curtivated for 3 days at 37 C. on a fish agar with systime and glucose. The antigenic substances were extracted from a rinsed, fathless, dry microbic mass.

The antigenic substances of B. tulescence were extracted by the following methods: I--entraction with 5% trychloracetic soid at 0 C. for 3 hours (Duaven method)(1); II--extraction by this came soid but for longer periods of time and at a higher temperature (modified Duaven method); III--extraction by phenol (A. P. Fonikov method)(1). The everage outcome of the antigenic substances were: for I-5.84%; for II--5.95%; for III--3.84%. The extracted substances were rinsed many times by reprecipitation of them with alcohol from water colutions. Subsidiary workin, a of the obtained preparations with chloroform (2) idicated an absence of any mechanical tinge of albumin in them. The extracted substance processed antigenic and allergenic properties, and also a serological activity, which gave a precipitating reaction in great dilutions on antitularence serums.

An attempt was made to descripted the untigons into separate components by means of heating for 1 hours in a boiling vater both, in the presence of 0.1% of acetic acid. Unfortunately the attempt was without results. Exchanging the hydrolysates with heated elloweform, we were able to extract an abundant residue from them, which dissolved well in

a mixture of equal parts of heated chloroform and sthyl alcohol. After removal of the chloroform entracted residue, one hydrolysate was worked with various quantities of ethyl alcohol.

As a result of the above working the antigons of the tularemia microbe were divided into 5 fractions: I-substances non-soluble in a minture of alcohol-chloroform (6.0-7.3%); II-residue from 4 volumes of alcohol (23.3-33.2%); III-residue from 10 volumes of alcohol (13.4-150); IV-substances extractable from the hydrolysate by chloroform and ether, plu substances which entered into a mixture of alcohol-chloroform during the obtaining of fraction I (33.9-37.8%); V- dry balance after removal of all the above fractions(11.8-18.2%). The antigens, extracted by verious conditions of trychloroacetic acid, were characterized by a similar content of separate fractions. The antigen obtained by the Konikova method differed from them somewhat.

A chemical analysis of the original antigens and products of their fractionation indicated that during hydrolysis with acetic acid the separation of the antigenic substances into chemically individual components (specific polysaccharides and albumin) does not take place (Table 1).

Each fraction, except the lipid (fraction IV), evidently, represents an intricate mixture of various elements of the antigen, or complexes containing carbohydrate as well as nitrous elements. Eucleic acids were detected in the composition of fraction II.

The studying of the quantitative composition of the carbohydrate and albuminous components of the original antigen, and their separate fractions, was done by chromatography on paper.

The composition of the monosaccharides, clears das a result of hydrolysis, was studied on single dimensional chromatogramms, takeing place in a mixture of n-butanch-acetic acid-water(40:10:50). As a

developer we used anilinghtalate in water-impregnated butanol. For the identification of the glucose and galacters we, besides this, used reactions with phorphydrazine, having in mind that the esazones of these monose are slightly different.

It was proven that all the antigens, irregardless of the method used for their entraction, contain quantitatively similar specific polysaccharides. Moselts of the chromatographic analysis of the polysaccharides of the antigens isolated by the Europen method, and the fractions II and III obtained from them, indicate that the carbohydrate components of the whole antigen and its fraction II are qualitatively of the same composition. They contain the following monocaccharides: galactose, arabinose, xylose, uronic acids and hexosamine. In the composition of the carbohydrate component of fraction III there were other saccharides detected, and namely: glucose, mannose, xylose and hexosamine(analyticly).

It is necessary to are that chromatographic analysis of the compocition of the polysaccharides of the original antigen disclosed no glucore or monnose. Judging by the intensiveness of the color of the stains on the chromatogramms, the basic mass of the polysabcharides of the original matigen consists of galactose and arabinose. The intensively dyeing stalks of these sugars on the chromatograms could have masked the weakly freing strains of the manage and glucore.

Chromatographic study of the albuminous components of the antigenic complexes, extracted by various methods from a virulent strain
of B. tularense, indicated that they have a qualificated by similar aninoacid content. Separation of the amenoacide of the hyprolyceton being
studied was done on one- and to-dimensional chromatograms. Mater impreparted buttened was used in a minture of n-buttened-acetic scid-nator

(40:10:50) as a solvent during the two-dimensional chromatography. In 0.2% solution of minhydrin was used in water-saturated butanole as a developer. The aminoacidic composition of the albumin of the antigen, extracted by the busyen method, is represented on the two-dimensional chromatogramm. L5 stains were located on the chromatogramm, of which one was unidentified. It is believed that it belongs to %,6-diamino-pymelic acid. Mistidine and lysine are located on this chromatogramm in one stain(spot). Their presence was indicated on the single-dimensional chromatogramms. The histidine, besides this, was detected by the Paul reaction. The presence of proline was indicated on a separate chromatogramm during the use of isatin. Consequently, the albuminous components of the antigen contain more than 17 aminoacids. In regard to the aminoacidic composition of the albuminous component, fraction II and III did not differ from each other.

The conducted studies established that the antigenic complex of the tularemia microbe, by its chemical nature, significantly differs from the full antigens of Buaven, from the bacteria of the intestinel-typhus group, studied to this time. Two complicated complexes-fractions II and III- enter into the composition of the tularemia antigen.

It was established that the latter complexes are separate components, firmly linked together. Thus, fraction II represents a stable complex, consisting of polysaccharides, albumin and nucleic acid, and fraction III-stable polysaccharides, albuminous complex. The albuminous components of the said fractions contain at least 17 similar aminoacids. The presence of qualitatively different polysaccharides in these fractions indicates; either two antigens, or one with various determinant groups.

Great interest is shown in the detection of an exceptionally high content of the lipid fraction in the tularemia antigen(approx. 40% from the original antigen), sharply differing from the analogical fractions of the Buaven full antigens, by chracteristics.

INST. EPID. MICHOB., imene H. F. GAMAIE. AMS., USSR., Entered for publication 14/may/55.

one table-copied two illustrations- not reproduced.

Literature

1. V. I. Tovernitski, Jell., 2, 37(1947); 10, 31 and 11, 82(1940).
2. IL G. Sevage, D. B. Zackren, A. Z. Smolens, J. Biol. Chem., 124, 425, (1938). 3. A. M. Belozercki, M. I. Prockuryskov, Prac. Vanuel for the Biochemistry of Plants, 1951. 4. S. M. Partridge, Nature, 164, 443 (49).
5. L. Elson, W. Morgan, Diochem. J., 27, 1824(1933).

ाऽस्, कं≢्र	• Antige	win(by cal-	rogen purine calculation)	rogen hexaced by calcu-	osamine	ucing sub-	onucleic acid	oxyribonuc- c wold **	ne acid (*	rus rus	con nitrogen	indicators	ì	
	ens obt	<u>-</u>	on the	0,9	11.0	20.7	# C i		i i	ho- rus 1.0	en 7,1	rs sr	Orig.	nical c
<u>-</u>	intigens obtained: E-Buaven, MS-Fodified Surven method, K-Konikov. Data of the substance ward determined only in fraction II. Fraction to Shm	<u>. </u>		9 0.9	0 10.9	7 20.2			· · · · · · · · ·	1.2	7.4	10 m	n racteristic	h ract
	B-Mare			0.8	9.6	20.1				0.9	5.9	1.4		Clemical ch racteristics of antigons
•	n, M3-	53.6		۰,	0	7.1				0.8	8.6	υ	I fr	
<u></u>	-l'odif mined	53.1		c	ပ	5.1				1.4	8.5	1113	I fraction	ıt i <i>g</i> ens
	ted Bu	35.4		o	0	6.9				0.6	5.7	K		and t
	ven m	35.5	1.7	1.3	16.8	27.9	10.5	5.2	1.5	1.7	8.7	Œ		heir s
	ethod.	35.9	1.8	1.1	13.4	28.5	10.9	5.1	1.5	1.5	3.5	F.B		and their segurate fract
<u> </u>	#10071 15071	!		1.1	14.0	26.7				1.9	æ. 5	×		
	Fraction tion to Shmidt and	49.2		1.6	20.3	25.5				0.6	9.5	ᇤ	0 III	fractions
· ~~~ <u>0</u>		49.7		1.6	20.2	25.0			· ; - · -	0.6	7.0		III fraction	(Jata 1
•	of the	53.3		1.5	19.6	25.4				0.6	Ŭ			(data expressed in / fron
<u> </u>	e phosp			0.2	2.8	5.6				1.2	<u>ڦِڙ</u>	*1	IV fraction	
	borus	, 		0.2	۵. 0	3.9				1.0	2.6	<u></u>	1	
	combin		·	ે.1	1.8	4.2				0.9	2.0	><	-	Try W
<u>a</u>	e phosphorus combinations according			0.6	7.7					ο 8	7.7	153	V fraction	fron dry weight of mrep.
	8 0000 7			0.9	10.9					0.8	·. ·s	<u> </u> ::	tion	of pre
	ling			0.5	5.8	25.9				0.3	7.5	2		ja.